

Heliannuol E. A Novel Bioactive Sesquiterpene of the Heliannane Family^{†,1}

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Received 15 January 1999; revised 23 April 1999; accepted 27 April 1999

Abstract: From the medium polar active fractions of leaf aqueous extracts of *Helianthus annuus* L. cv. SH-222, we have isolated heliannuol E (1) which contains a previously unknown six-membered ring containing bicyclic skeleton. The structural elucidation is based on extensive spectral studies, including ¹H-¹H COSY, ¹H-¹³C HETCOR, and NOE difference experiments. © 1999 Elsevier Science Ltd. All rights reserved.

The discovery of new allelochemicals from plants or microbes has attracted much attention in the last 20 years. Allelochemicals have been implicated as biocommunicators, ² and are potential sources of new structural types of herbicides with new modes of action which may be less harmful than those presently used in agriculture. ³ It is becoming increasingly important to develop premixes of herbicides employing different modes of action, so as to get both short-term efficacy and long-term control of a given weed spectrum. Allelopathy, ⁴ an emerging branch of applied sciences which studies biochemical plant-plant and plant-microorganism interactions, may help in providing new approaches to integral weed control management by providing, for instance, crop varieties having greater ability to smother weeds, generations of natural phytotoxins as herbicides and of synthetic derivatives of natural products as herbicides.

Sunflowers are well-known for producing a variety of secondary metabolites, most commonly terpenoids⁵ and phenolics⁶ possessing a wide variety of biological properties⁷ including allelopathic activity.⁸ In our screening of crude extracts for phytotoxic allelopathic activity⁹ that can be used as natural herbicide templates,¹⁰ the medium polar active fractions of leaf aqueous extract of two varieties of cultivar sunflowers (*Helianthus annuus* L.) cvs. SH-222[®] and VYP[®] displayed a potent profile of activity particularly against dicotyledons. Bioassay-guided

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fractionation of this extract afforded, together with the known heliannuols A-D¹¹ the novel heliannuol E (1) which contain a previously unknown six-membered ring containing bicyclic skeleton.

The extraction of the fresh leaf aqueous extract of *H. annuus* L. cv. SH-222[®] with dichloromethane gave material which was fractionated by chromatography on silica gel using hexane-ethyl acetate mixtures of increasing polarity. Plant material was collected during the third plant development stage⁹ (plant 1.2 m tall with flowers, 1 month before harvest). Bioassay-guided fractionation of this extract afforded heliannuol E (1).

Table 1. ¹H NMR data (400 MHz, *J*

values in Hz) for I (CDCl ₃)			
H	6		
3	6.65 s		
6	6.47 s		
7	5.95 ddd		
	$(J_{7,14}=10.1; J_{7,8\alpha}=6.2; J_{7,14}=17.0)$		
8α	3.46 ddddd		
	$(J_{7,8\alpha}=6.2; J_{8\alpha,9\alpha}=5.5; J_{8\alpha,9\beta}=2.2;$		
	$J_{8\alpha,14}=J_{8\alpha,14}=1.5$)		
9α	1.89 ddd		
	$(J_{\mathbf{z}_{\alpha},9\alpha}=5.5; J_{9\alpha,9\beta}=13; J_{9\alpha,10}=10.1)$		
9β	1.85 ddd		
	$(J_{8\alpha,9\beta}=2.2; J_{9\alpha,9\beta}=13; J_{9\beta,10}=3)$		
10	3.72 dd		
	$(J_{9\alpha,10}=10.1;J_{9\beta,10}=3)$		
12	1. 28° s		
13	1.23° s		
14	5.01 ddd		
	$(J_{7,14}=10.1; J_{14,14}=J_{8a,14}=1.5)$		
14'	4.89 ddd		
	$(J_{7,14}=17.0;J_{14,14}=J_{8\kappa,14}=1.5)$		
15	2.18 s		

^{*} These values may be interchanged.

Figure 1. Heliannuol E (1)

Heliannuol E (1) showed a HRMS spectrum with a molecular ion at m/z 248.1420, corresponding to the molecular formula C₁₅H₂₀O₃. Both ¹H NMR (Table 1) and ¹³C NMR spectra¹² indicated that it had a heliannuol skeleton: δ 6.65, 6.47, 2.18 in the ¹H NMR spectrum and δ 124.5, 148.2, 118.5, 120.6, 147.5, 115.8 in the ¹³C NMR spectrum. ¹H NMR spectra indicated the presence of a vinylic moiety, similar to that shown by heliannuol C (4)¹¹ [H-14(δ 5.01), H-14'(δ 4.89), H-7(δ 5.95)]. Study of 2D COSY and 2D HETCOR spectra confirmed the same oxygenation pattern as in 4: H-7 is coupled with H-8 (\ddot 3.46), H-8 with H- 9α (δ 1.89), H-9 β (δ 1.85) and H-9 α , H-9 β with H-10 (δ 3.72). Coupling constants suggested a six-membered ring, with H-8a and H-9b at equatorial positions and H-9α and H-10β at axial ones. The chemical shift observed for the signal corresponding to C-11 (& 71.1) is in good agreement with an ether function at C-10, as in heliannuol B (3)¹¹ (C-11, δ 72.4) and D (5) (C-11, δ 72.8) instead of an ether function attached at C-11 as in heliannuol A (2) (C-11, δ 79.7) and C (4)¹¹ (C-11, δ 79.7). Consequently, the structure was assigned as represented in Figure 1 and constitutes the first isolated heliannuol with a six-membered ring. The stereochemistry was assigned on the basis of a NOE experiment (figure 3),

Figure 2. Heliannuols A-D

where effects between H-10 and H-7; H-10 and H-9 β ; as well as between H-8 and H9 α were observed. Both, the stereochemistry and the presence of a six-membered ring are corroborated with the good agreement observed between the angles found for the most stable conformation and the experimental coupling constants (Table 2).

Table 2. Observed coupling constants vs **Φ** obtained for the most stable conformer

Protons	Calculated 4	Observed J (Hz)
8α-9α	41.5	5.5
8α-9β	75.2	2.2
9α-10β	179.6	10.1
9β-10β	64.9	3.0

Figure 3. Observed NOEs for the most stable conformer of 1 using PM3 calculations. 13

This compound is of particular interest since it is the first member of a new heliannuol skeleton with potential allelopathic activity. Based on its bioassay results, ¹⁴ it is likely to be significantly involved in the allelopathic action of cultivar sunflowers. Its biogenesis may proceed through a bisabolene-type precursor as proposed for the recently reported heliannuols. ¹¹ As for 4, its biosynthesis cannot be easily accommodated by the isoprene rule, but may involve a rearrangement via a phenonium ion to afford the six-membered heterocyclic ring.

Acknowledgments. This research has been supported by the Dirección General de Investigación Científica y Técnica, Spain (DGICYT; Project No. PB95-1231). We thanks Dr. Albert García de Luján y Gil de Bernabé and Mr. Miguel Lara, CIFA, Rancho de la Merced, Agricultural Research Station, Junta de Andalucía, Jerez, Spain for providing plant material.

REFERENCES AND NOTES

- † This paper is affectionately dedicated to Professor Francisco Rodríguez-Luís on the occasion of his 60th birthday, wishing him the best in life and in chemistry.
- Part 12 in the series "Allelopathic studies in cultivar species" for part 11 see: Macías, F.A.; Varela, R. M.; Torres, A. and Molinillo, J.M.G. Tetrahedron Lett. 1998, 39, 427.
- (a) Fischer, N.H. In The Science of Allelopathy; Putnam, A.R., Tang, C.S., Eds.; John Wiley & Sons: New York, USA,1986; Chapt.12, pp 203-218.(b) Macías, F.A.; Oliva, R.M.; Simonet, A.M; Galindo, J.C.G. In Allelopathy in Rice; Oloffdoter, M. Ed.; I.R.R.I. Press: Los Baños, Philippines, 1998; Chapt. 7, pp 69-79.(c) Harborne, J. B. Nat. Prod. Report. 1993, 10, 327. (d) Harborne, J. B. Nat. Prod. Report. 1997, 14, 83.
- (a) Duke, S.O. Reviews of Weed Science 1986, 2, 15. (b) Macías, F.A. In Allelopathy: Organisms, Processes, and Applications; Inderjit, Dakshini, K.M.M., Einhellig, F.A., Eds.; ACS Symposium Series, 582: Washington, D.C., USA, 1995; Chapt. 23, pp 310. (c) Rice, E.L. Biological Control of Weeds and Plant Diseases, University of Oklahoma Press: Norman. USA, 1995.
- 4. (a) Rice, E.L. Allelopathy; Second Edition; Academic Press: New York, USA, 1984. (b) Allelopathy is officially defined as: The science that studies any process involving, mainly, secondary metabolites produced by plants, algae, bacteria, and fungi that influence the growth and development of agricultural and biological systems, including positive and negative effects, IAS International Allelopathy Society Constitution (1996), First Word Congress on Allelopathy. A Science for the Future, September, Cádiz, Spain.
- (a) Spring, O; Albert, K.; Gradmann, W. Phytochemistry 1981, 20, 1883. (b) Beale, M.H.; Bearder, J.R., MacMillan, J.; Matsuo, A.; Phinney, B.O. Phytochemistry 1983, 22, 875. (c) Pyrek, J. S. J. Nat. Prod. 1984, 47, 822. (d) Spring, O.; Rodon, U.; Macías, F.A. Phytochemistry 1992, 31, 1541. (e) Alfatafta, A.A.; Mullin, C.A. Phytochemistry 1992, 31, 4109. (f) Macías, F.A.; Varela, R. M.; Torres, A.; Molinillo, J.M.G. Phytochemistry 1993, 34, 669. (g) Bushman, H.; Spring, O. Phytochemistry, 1995, 39, 367. (h) Macías, F.A.; Torres, A.; Molinillo, J.M.G.; Varela, R. M.; Castellano, D. Phytochemistry 1996, 43, 1205.
- (a) Rieseberg, L.H.; Soltis, D.E.; Arnold, D. Am. J. Botany 1987, 74, 224. (b) Schilling, E.E. Biochem. Syst. Ecol. 1983, 11, 341. (c) Mazza, G.; Gao, L. Phytochemistry 1994, 35, 237. (d) Macías, F.A.; Molinillo, J.M.G.; Torres, A.; Varela, R.M.; Castellano, D. Phytochemistry 1997, 45, 683.
- (a) Ray, S.B., Guruprasad, K.N.; Laloraya, M.M. J. Exp. Botany 1980, 31, 1651. (b) Spring, O; Benz, A.; Faust, V. J. Plant Dis. Protect. 1991, 98, 597.
- (a) Saggese, E. J.; Foglia, T. A.; Leather, G; Thompson, M.P.; Bills, D.D.; Hoagland, P.D. In *The Chemistry of Allelopathy* Thompson A.C. Ed.; ACS Symposium Series, 268: Washington, D.C., USA, 1985; Chapt. 7, pp 99-112.
 (b) Leather, G.R. *Plant Soil* 1987, 98, 17. (c) Macías, F.A.; Varela, R. M.; Torres, A.; Molinillo, J.M.G. In *Allelopathy in Pests Management for Sustainable Agriculture* Narwal, S.S.; Tauro, P.; Eds.; Scientific Publishers: Jodhpur, India, 1996; Chapt. 4, pp 77-92.
- Macías, F.A.; Varela, R.M.; Torres, A.; Molinillo, J.M.G. In Principles and Practices in Chemical Ecology Inderjit;
 Dakshini, K.M.M.; Foy, L; Eds.; CRC Press. LLC: Boca Raton, FL, USA, 1999; Chapt. 27, pp 531-550.
- Macías, F.A.; Castellano, D.; Oliva, R. M.; Cross, P; Torres, A. In Proceedings of Weeds. The 1997 Brighton Crop Protection Conference, Copping L.G., Ed.; British Crop Protection Council: Farnham, UK, 1997; Vol 1; pp 33-38.
- (a) Macías, F.A.; Varela, R. M.; Torres, A.; Molinillo, J.M.G.; Fronczek, F.R. *Tetrahedron Lett.* 1993, 34, 1999-2002.
 (b) Macías, F.A.; Molinillo, J.M.G.; Varela, R. M.; Torres, A. and Fronczek, F.R. *J. Org. Chem.* 1994, 59, 8261-8266.
- 12. Yellow oil; $[\alpha]^{D}_{25} = -68.6$ (CHCl₃, c=0.1); IR ν_{max} (meat, KBr) cm⁻¹:3423 (hydroxyl group); 1632 (C=C); 1195 (ether);

 1H NMR is listed in Table 1; ¹³C NMR δ : 124.5 (s, C-1); 148.2 (s, C-2); 118.5 (d, C-3); 120.6 (s, C-4); 147.5 (s, C-5); 115.8 (d, C-6); 142.1 (d, C-7); 37.9 (d, C-8); 27.5 (t, C-9); 78.1 (d, C-10); 71.1 (s, C-11); 25.1 (q, C-12); 24.2 (q, C-13); 113.1 (d, C-14), 15.7 (q, C-15).; EIMS, m/z (rel. int.): 248 [M]* (86.5), 230 [M H₂O]* (58.5), 23 [M-CH₃]*(9.5), 215 [M-H₂O-CH₃]* (70), HREIMS calcd for C₁₃H₂₀O₃ 248.1412, found 248.1420.
- 13. Stewart, J.J.P. *J. Comput. Chem.* **1989**, 10, 209. Molecular orbital calculations were carried out using PM3 Hamiltonian as implemented in MOPAC 6.0. Geometries were fully optimized using the PRECISE keyword.
- 14. Varela, R.M. "Allelochemicals of the variety cultivar sunflower SH-222. Heliannuols and Heliespirones two novel families of sesquiterpenes" Doctoral Thesis. University of Cadiz, Spain. 1996.